

**In the Specification:**

Please replace the paragraph beginning at page 5, line 31, with the following:

*A1*  
--As used herein, the term "single-chain antibody" refers to a polypeptide comprising a V<sub>H</sub> domain and a V<sub>L</sub> domain in polypeptide linkage, generally linked via a spacer peptide (e.g., [Gly-Gly-Gly-Gly-Ser]<sub>x</sub>; SEQ ID NO:1), and which may comprise additional amino acid sequences at the amino- and/or carboxy-termini. For example, a single-chain antibody may comprise a tether segment for linking to the encoding polynucleotide. As an example, a scFv is a single-chain antibody. Single-chain antibodies are generally proteins consisting of one or more polypeptide segments of at least 10 contiguous amino acids substantially encoded by genes of the immunoglobulin superfamily (e.g., see The Immunoglobulin Gene Superfamily, A. F. Williams and A. N. Barclay, in Immunoglobulin Genes, T. Honjo, F. W. Alt, and T. H. Rabbitts, eds., (1989) Academic Press: San Diego, Calif., pp. 361-387, which is incorporated herein by reference), most frequently encoded by a rodent, non-human primate, avian, porcine, bovine, ovine, goat, or human heavy chain or light chain gene sequence. A functional single-chain antibody generally contains a sufficient portion of an immunoglobulin superfamily gene product so as to retain the property of binding to a specific target molecule, typically a receptor or antigen (epitope). Techniques for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce antibodies for use in this invention.--

Please replace the paragraph beginning at page 31, line 30, with the following:

*A2*  
--The H11 library described above was constructed from a 50 kb human P1 (P1 clone 876h9, Genbank accession AC004039), containing the Interleukin-4, Interleukin-13, and kinesin-like protein-3 genes from 5q31. 20 µg P1 DNA was purified by standard method (Qiagen) (Collins *et al.*, *Proc. Natl. Acad. Sci USA* 95:8703-8708,

1998) and was randomly fragmented with decreasing concentrations of DNase I (10 units / ml) in 10 mM Tris pH 7.0 / 10 mM MnCl<sub>2</sub> for 8 minutes at 15°C, extracted and precipitated. Fragments were blunted with 5 units/μg T4 polymerase for 30 min at 12°C, extracted and precipitated. Linkers containing a Sfi-1 restriction site (Link1 5'-AGCGGCCGCAGGCCATGGAGGCC-3' (SEQ ID NO:2), Link2 5'-GCCCTCCATGGCCTGCAGGCCGCT-3' (SEQ ID NO:3)) were ligated to target DNA with 400 units T4 DNA ligase for 2 hours at room temperature. The resulting product was electrophoresed on a 2.0% agarose gel and the size range of 100-300 bp was collected and eluted from NA-45 DEAE paper (Schleicher and Schuell, Keene, NH) 100 ng of the linker-ligated product was used as template in PCR with a nested primer LP5 (5'-GCGGCCGCAGGCCATGGA-3'; SEQ ID NO:4) with 2.5 units Pfu Polymerase/2.5 units panoTaq for 30 cycles (94°C x 1 min, 55°C x 1 min, 72°C x 1 min). The PCR products were digested with Sfi-1 and gel purified. A positive control phage displaying the 3' exon of the IL-4 cDNA (490-612 bp) was also constructed (Yokota *et al.*, *Proc. Natl. Acad. Sci USA* 83:5894-5898, 1986).--

*Q2*  
*Cont.*

Please replace the paragraph beginning at page 32, line 14, with the following:

*Q3*

--A phage display vector, pORF-1, was engineered for gene fragment phage display. It is a pHEN-1 (Hoogenboom *et al.*, *Nucl. Acid Res.* 19:4133-4137, 1991) based vector that contains a pelB leader sequence, a 5' hexahistidine tag and a non-religatable Sfi-1 insert cloning site which is upstream and contiguous with the M13 gene III and a 3' myc epitope tag. pORF-1 was constructed by two rounds of template mutagenesis of pHEN-1 vector with primers (NSFI 5'-GCGGCCAGCCGGCGATGCC CCAGCACCATCACCATCACGGGGCCATGGTGCAGCTGCAGG-3' (SEQ ID NO:5); SUP 5'-TCACGGGGCCATGGGGCCCAGGCCTCAGTCGATCGACACGG CCTCCACGGCCGCAGAACAA-3' (SEQ ID NO:6)) (Kunkel *et al.* *J. Biol. Chem.* 263:14784-14789, 1988). The base vector contained an out-of-frame 1 kb stuffer fragment. Sfi-1 digested insert was ligated into the digested vector and optimized